Biological Control of Water Snails by Dragonfly Nymphs

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Abstract The predatory potential of dragonfly nymphs was studied on water snails (Indoplanorbis exustus and Lymnaea luteola) and their larval stages under laboratory conditions. The water snails (snail’s cocoons, young snails and adult snails etc.) and dragonfly nymphs were collected from stagnant water resources of protected areas and brought to laboratory, maintained with aquatic weeds and mulberry leaves in the artificial cement tank. The collected water bodies were used in the requisite quantity in glass containers of 6 litre water capacity each with snails, aquatic weeds and dragonfly nymphs for trial groups while the container of control group was kept without dragonfly nymphs. Observation recorded from 0, 3, 6, 12, 24, 48, 72 and 96 hrs of nymphs entry while feeding activity of nymphs was judged by counting the remaining numbers of snails and their larval stages in the different containers. The pond water quality analysis was envisaged, average dissolve oxygen (DO 0.4—0.7 ppm), hydrogen ion concentration (pH 6.7-8.7) and total dissolve solids (TDS 71-84 ppm) during the entire period of experiment. Remarkable reduction (82-97 %) in the cocoons and young snail population were noticed whereas 18-40% mortality have also encountered in the snails of control group. The predacious nature and potentials of the dragonfly nymphs may be promoted as one of the appropriate biological control agent to overcome the snail borne parasitic diseases.

Keywords Biological Control, Water Snails, Nymphs, Dragonfly

1. Introduction

The dragonflies (Palaeophlebia) are useful predacious insect and their nymphs grow in the fresh water resources like rainy season ditches, river pools, ponds and marshy lands etc. Similarly the water snails (Indoplanorbis exustus and Lymnaea luteola) are also habitat to the stagnant water resources [1, 2]. The fluke eggs usually passed in the faeces of domestic as well as wild herbivores and under suitable conditions of moisture and warmth larva called miracidia hatches and infect the water snails that act as intermediate host [15]. Expansions of parasitic diseases are worldwide owing to the human rehabilitation and indiscriminate uses of natural resources. Fluke borne diseases are creating an alarming situation to sustenance of domestic and wild ruminants with the loss of agility due to apoptosis in the infected host cells that decrease the immunocompetency resulting untimely demises [5]. Frequent breeding of snails around the human community and their livestock occurs due to lift irrigation, pool of water and conventional water logging system in the rural areas. Thus it became major problem to the animal industry growth as worldwide 300 million bovines found annually exposed with the fluke parasites causing economic losses amounting more than 3.0 Billion US$ [7].

Inadequate uses of chemotherapeutics in practices with the reliance on chemical control have commenced harmful circumstances to the ecosystem with residual toxicity and resistance against the parasitic infections in animals [4, 8].

Withstanding the facts of drug resistance, the biological control measures are reasonably appropriate as they are eco-friendly and beneficial for sustenance of aquatic flora and fauna. There are several biological control agents found in the fresh water resources like larvivorous fishes, water bugs, Coleopteran aquatic larvae, Crayfish, Cyclopes and Echinostome snails etc., those naturally helps in controlling the population of snails though they have their own barriers and capacity [9]. However, the sustainable alternative needs to have distinct advantages and ability to kill the target species with easy applications, less expensive and non-pathogenic to biotic community [6]. Such qualities have together with the dragonflies and their larval stages as they are entomophagus in nature and frequently feed upon snails with longevity to spend their three- forth life span (1-3 years) in the aquatic habitat [10]. Michelson [11], Somasunderao [14], Bali et al. [3] have used dragonfly nymphs as biological control agent to overcome the schistosome bearing snails while Singh et al. [13] also used for control of malaria vector.

In the present study, an attempt was made to assess the predatory potentials of dragonfly nymphs as biological control of water snails.

2. Materials and Methods

A series of laboratory based experimental trials on dragonfly nymphs were conducted since 2010, aiming with daily predation rate, predatory preferences on snail cocoons,
small and large size snails, selective predation on different species of snail’s and their dietary requirements etc.

The water snails (*Indoplanorbis exustus* and *Lymnaea luteola*) with their larval stages and dragonfly nymphs were collected from stagnant water resources of protected areas and maintained into artificially made cement tank with aquatic weeds. Further they were kept in different classified containers with aquatic weeds, lotus and mulberry leaves as the snails are frequently feed upon the mulberry leaves and shelter their cocoons under the surface of leaves of Lotus (*Nilumb* *nucifera*) and water chestnuts (*Trapa natans*). After repeated outcomes and encouraging observations, the laboratory experiment was scheduled in the glass containers (12”x12”x 6”) with 5-liters of stagnant water. The predatory potential of the dragonflies’ nymphs were assessed on the basis of snails feeding status with their larval stages in each container by dragonfly nymphs. The experiment was designed with seven groups: I - Cocoons (100), II- Young snails (100), III- Adult snails (100), IV- Cocoons (100) and Young snails (100), V- Young snails (100) and Adult snails (100); VI- Cocoons (100), Young snails(100) and Adult snails (100) and VII- Cocoons (100), Young snails(100) and Adult snails (100) without nymphs of dragonfly (Control).

Five nymphs of dragonfly were released in the containers of 1-6 groups and the feeding behaviour and their activities were recorded between 0-96 hrs (Table 1) while the water quality analysis (Table 2) was also performed at 0, 24, 72 and after a week intervals for Hydrogen ion concentration of the water (pH), dissolve oxygen (DO) and total dissolve solids (TDS) by commercially available water quality analysis kits (Aqua-Check Pvt. Ltd. India). However, for snail’s feed, the fresh mulberry leaves were used on alternate days and the faecal material of nymphs or dead snail’s shells were routinely discarded. For appropriate counting of dead and live snails, the method of Agrawal [1] was used as dead snails sink frequently in the jar water.

### Table 1. Feeding Behavior of Dragonfly Nymphs on larval stages of snails

<table>
<thead>
<tr>
<th>Observations</th>
<th>I</th>
<th>II</th>
<th>III</th>
<th>IV</th>
<th>V</th>
<th>VI</th>
<th>Control VII</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Temp °C</td>
<td>C</td>
<td>YS</td>
<td>AS</td>
<td>C</td>
<td>YS</td>
<td>AS</td>
</tr>
<tr>
<td>Zero hrs</td>
<td>28.5</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>After 3 Hrs</td>
<td>28.5</td>
<td>90</td>
<td>97</td>
<td>98</td>
<td>90</td>
<td>97</td>
<td>98</td>
</tr>
<tr>
<td>After 6 Hrs</td>
<td>29.0</td>
<td>85</td>
<td>90</td>
<td>98</td>
<td>85</td>
<td>82</td>
<td>89</td>
</tr>
<tr>
<td>After12 Hrs</td>
<td>29.2</td>
<td>75</td>
<td>71</td>
<td>95</td>
<td>75</td>
<td>64</td>
<td>74</td>
</tr>
<tr>
<td>After 24 Hrs</td>
<td>29.2</td>
<td>54</td>
<td>41</td>
<td>91</td>
<td>67</td>
<td>42</td>
<td>45</td>
</tr>
<tr>
<td>After 48 Hrs</td>
<td>29.0</td>
<td>15</td>
<td>28</td>
<td>90</td>
<td>41</td>
<td>32</td>
<td>18</td>
</tr>
<tr>
<td>After 72 Hrs</td>
<td>29.0</td>
<td>03</td>
<td>12</td>
<td>85</td>
<td>06</td>
<td>14</td>
<td>00</td>
</tr>
<tr>
<td>After 96 Hrs</td>
<td>28.5</td>
<td>00</td>
<td>00</td>
<td>78</td>
<td>00</td>
<td>00</td>
<td>00</td>
</tr>
<tr>
<td>Left alive</td>
<td>-</td>
<td>00</td>
<td>00</td>
<td>78</td>
<td>00</td>
<td>00</td>
<td>00</td>
</tr>
</tbody>
</table>

C = Cocoons, YS=Young Snail, AS = Adult Snails

### Table 2. Quality Analysis of used Pond Water

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Temp °C</th>
<th>Observation Time</th>
<th>Normal Values</th>
<th>I</th>
<th>II</th>
<th>III</th>
<th>IV</th>
<th>V</th>
<th>VI</th>
<th>VII</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH (Hydrogen Ion concentration)</td>
<td>28.5</td>
<td>Zero day</td>
<td>7.1</td>
<td>6.8</td>
<td>6.5</td>
<td>0.5</td>
<td>0.7</td>
<td>0.6</td>
<td>0.5</td>
<td>0.5</td>
</tr>
<tr>
<td></td>
<td>28.5</td>
<td>After 24 hrs</td>
<td>6.9</td>
<td>6.6</td>
<td>0.6</td>
<td>0.5</td>
<td>0.5</td>
<td>0.6</td>
<td>0.6</td>
<td>0.4</td>
</tr>
<tr>
<td></td>
<td>29.0</td>
<td>After 72 hrs</td>
<td>6.9</td>
<td>6.8</td>
<td>0.6</td>
<td>0.5</td>
<td>0.6</td>
<td>0.6</td>
<td>0.5</td>
<td>0.5</td>
</tr>
<tr>
<td></td>
<td>29.2</td>
<td>After 96 hrs</td>
<td>6.9</td>
<td>6.8</td>
<td>0.5</td>
<td>0.5</td>
<td>0.5</td>
<td>0.4</td>
<td>0.5</td>
<td>0.5</td>
</tr>
<tr>
<td>DO/ppm (Dissolve Oxygen)</td>
<td>28.2</td>
<td>Zero day</td>
<td>0.5</td>
<td>0.6</td>
<td>0.5</td>
<td>0.4</td>
<td>0.5</td>
<td>0.5</td>
<td>0.5</td>
<td>0.4</td>
</tr>
<tr>
<td></td>
<td>29.0</td>
<td>After 24 hrs</td>
<td>0.5</td>
<td>0.5</td>
<td>0.5</td>
<td>0.4</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0.4</td>
</tr>
<tr>
<td></td>
<td>29.0</td>
<td>After 72 hrs</td>
<td>0.5</td>
<td>0.5</td>
<td>0.5</td>
<td>0.4</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0.5</td>
</tr>
<tr>
<td></td>
<td>28.5</td>
<td>After 96 hrs</td>
<td>0.5</td>
<td>0.5</td>
<td>0.5</td>
<td>0.4</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
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<tr>
<td>TDS/ppm (Total Dissolve Solids)</td>
<td>29.5</td>
<td>Zero day</td>
<td>80-90</td>
<td>78</td>
<td>75</td>
<td>72</td>
<td>78</td>
<td>72</td>
<td>76</td>
<td>75</td>
</tr>
<tr>
<td></td>
<td>29.5</td>
<td>After 24 hrs</td>
<td>74</td>
<td>78</td>
<td>76</td>
<td>78</td>
<td>78</td>
<td>76</td>
<td>76</td>
<td>76</td>
</tr>
<tr>
<td></td>
<td>29.0</td>
<td>After 72 hrs</td>
<td>75</td>
<td>76</td>
<td>72</td>
<td>71</td>
<td>74</td>
<td>84</td>
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</tr>
<tr>
<td></td>
<td>29.2</td>
<td>After 96 hrs</td>
<td>78</td>
<td>78</td>
<td>76</td>
<td>81</td>
<td>79</td>
<td>78</td>
<td>84</td>
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</tbody>
</table>
3. Results and Discussion

The dragonfly nymphs have natural predation upon snails. They frequently feed on cocoons as well as young snails of *L. luteola* and *I. exustus* (82-97%) in I-VI groups while unable to prey on adult snails (Table 1 Fig 1). It was also noticed that nymphs are mostly prefers bottom surface where only adult snails resides and due to their big size, these nymphs are found to be incapable to neutralize the adult snails (Fig 1b). The cocoons of the snails reside under the surface of the lotus leaves and usually remain safe (Fig 1a). Survival of the larval stages of the snails in the natural water resources depends upon the ample quantity of aquatic weeds and leaves to escape from predatory insects [2] whereas in the present experiment, glass containers (12”x12”x 6”) with 5-liters of stagnant water were found to be inadequate, hence the nymphs could easily find and predate upon the larval stages of the snails. Bali et al. [3] and Somasunderao [14] have also tried the aquatic insect in the laboratory conditions for controlling the snail borne diseases of live stock with unidentified species of the insects.

During the experiment, it was found that the cocoons and young snails (Group I and II) were fed upon by the nymphs within 96 hours of their entry in the containers. However, three nymphs were found dead on 4th day of experiment and remaining live nymphs have utilized the corpse as feeding material. Wissinger [16] have also observed similar incidence of cannibalism with high intensity of competition and predation during insufficient food materials. On the other hand, in the containers of Group III, IV, V and VI the nymphs were found to be alive and motile. It might be owing to sufficient quantity of young snails and cocoons (200) instead of group I and II. Thus, the experiment has shown that the 100 young snails are sufficient food material for five nymphs for 3-4 days survival and may be useful to design the mass fluke eradication programme in near future. However, in the control group 18-40% natural mortality of snails was encountered within week whereas Agrawal et al. [2] have reported 19-39 % mortality in snails within 30 days in artificially made water cement tank.

The biological control agent used in the present investigation to overcome the fresh water snails (*Indoplanorbis exustus* and *Lymnaea luteola*) have also found to be eco-friendly and convenient to overcome the bio-degradation of water resources as no harmful effect has been observed in the water quality analysis (Table 2).

Hofkin et al. [12] have tried Crayfish (*Procambrus Clarkii*) to overcome the schistosome-transmitting snails in the laboratory and temporary man-made ponds in Kenya with encouraging results. In addition to that, the German scientists Xinhua and Meyer-Rochow [17] have also investigated the morphological and behavioral adaptation of the aquatic larvae of *Aquatica lei* (*Colepota; Lampridae*) to prey upon fresh water snails that serve as intermediate host for liver fluke. They have carried out the series of laboratory based predation experiment and found the larvae of *Aquatica lei* (*Colepota; Lampridae*) revealed well developed pygypodia and specialized mouth parts to attack and overpower the water snails while they have wide dietary spectrum and can also feed on dead snails with similar feeding pattern of nymphs of dragonflies as reported in the present experiment.

The findings of the present study may be utilised as biological control agents to overcome the harmful pathogens for better health status of wild and domestic animals including humans.

Figure 1. A: Snails and their Cocoon on Lotus leaf B: Dragonfly Nymphs defending with adult snails C: Nymph feeding on young snails D: Nymphs fighting each other
4. Conclusions

Fluke borne diseases are major threats to the livestock industry as well as wild ruminants though biological control agents have capacity to control them. The assessment of the present study is suggested for the initiation of breeding programmes for the development of huge number of dragonfly nymphs and their introduction in different ponds, ditches and marshy lands that may be succeeded to check the population of water snails.

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