

Anabolic Steroid Treatment Induces Cardiac Autonomic Dysfunction in Rats: Time-Course of Heart Rate Variability

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Abstract The aim of this study was to investigate time-course of cardiac autonomic activity in rats treated with nandrolonedecanoate (DECA). Twenty male Wistar rats received weekly 10 mg.Kg⁻¹ of DECA (DG) or vehicle (CG) during 8 weeks. To heart rate variability analysis SDNN (standard deviation of RR intervals), RMSSD (square root of the mean squared differences of successive RR intervals), pNN5 (percentage of successive RR interval differences greater than 5ms) and Poincaré analysis was applied in time-domain. Additionally, power spectral was decomposed into high (HF: 0.8-2.5Hz) and low-frequency (LF: 0.2-0.85Hz). For the HF component, differences were found between DG and CG during 4th, 6th and 8th weeks. Progressive increase in the LF-HF ratio was found in DG, resulting in significant differences between groups in 4th, 6th, 7th and 8th weeks. DG showed decreased from 24.5±7.2ms² to 3.9±2.55ms² in HF corresponding (83.9% of alteration). Power spectral density function for HF corresponded to 26.3±8.3% and 26.5±4.1% in the 1st and 8th weeks (P=NS) for CG, whereas the DG showed reduced significantly from 26.4±3.6% (1st) to 18.99±2.7% (8th week). Cardiac autonomic dysfunction may constitute an early consequence of DECA administration and an important marker of arrhythmia vulnerability and sudden death identification.

Keywords Anabolic-Androgenic Steroid, Autonomic Nervous System, Heart Rate Variability, Spectral Analysis, Nandrolonedecanoate, Poincaré Analysis

1. Introduction

Testosterone is an androgenic-anabolic hormone synthesized in the Leydig cells in men and theca cells in women. Testosterone and other androgenic-anabolic steroids (AAS) stimulate the commitment of pluripotent mesenchymal stem cell toward myogenic lineage rather than adipogenic lineage, hence increase the size of both type I and type II muscle fibers [1, 2]. AAS are synthetic derivatives of testosterone, which were developed in order to enhance the anabolic and, otherwise, to attenuate the androgenic effects of testosterone [3]. Given the significant anabolic effect and the associated tissue building properties, Nandrolonedecanoate (DECA) is one of the most often AAS consumed around the world by athletes and non-athletes [4] to improve the physical performance related to muscular mass

and strength [5, 6]. The physiological action of AAS may be summarized as a migration of a steroid-receptor complex to the nucleus, where the transcription of genes into mRNA promotes protein synthesis [7, 8]. Nonetheless, the use of high-doses of AAS during the last years has been attributed as the main cause of several cardiovascular disorders, including arterial hypertension [9, 10], lipid profile abnormalities [11], heart failure [12], hypertrophic cardiomyopathy [13], arrhythmia [14], and sudden death [15, 16], generating a serious public health problem [17].

Nowadays, the analysis of heart rate variability (HRV) has been widely used as an indirect assessment of cardiac autonomic alterations, associated to immune dysfunction, inflammation, as well as a large range of cardiovascular diseases and sudden death [18-21]. Considering the decreased vagal function as a key factor for all the major risk factors for cardiovascular diseases [21], the HRV analysis may provide a noninvasive method for estimating the sympatho-vagal balance [20, 22], providing independent prognostic information about ventricular arrhythmia [23-25]. Accordingly, both the power spectral [20, 23] and the

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Poincaré[26] analysis of HRV has been applied to identify cardiac electrical instability. Nevertheless, while marked reductions in the parasympathetic activity was associated to cardiac autonomic impairment after treating with DECA [25], the role of the autonomic nervous system and AAS effects, showing the autonomic early possible effects induced by chronic high doses of AAS remains unknown. Thus, the purposes of this study were: 1 - to investigate “in vivo” the time-course of cardiac autonomic dysfunction in rats treated with DECA, using time- and frequency-domain HRV analysis; and 2 – evaluate a nonlinear method to analyze HRV in this experimental model.

2. Material and Methods

Experimental Animals

Twenty male Wistar rats were kept at 25 ± 2 °C with free access to rat chow and water ad lib following the guidelines of the institutional animal care and use committee. Animals were randomly allocated into two experimental groups: Control Group (CG, n=10) and DECA Group (DG, n=10). All animals received humane care in compliance with the “Principles of Laboratory Animal Care” formulated by the National Society for Medical Research and the “Guide for the Care and Use of Laboratory Animals” published by the National Institutes of Health (NIH publication 85-23, revised 1985).

During 8 weeks DG received 10 mg.Kg^{-1} of intramuscular DECA (DecaDurabolin, Organon®; gluteus medium) weekly and CG received the same volume of vehicle, composed of peanut oil with benzyl alcohol (90:10, V/V) [27, 28]. This dose was calculated from the therapeutic dose of nandrolone decanoate (about 1 mg.Kg^{-1} each 3 weeks). The dose used in the study presents values considered supraphysiological. Reports of use by elite athletes had shown doses between 350-600mg weekly, which corresponds to a rate of approximately 5-7mg/kg (NATIONAL INSTITUTE ON DRUG ABUSE, 2013).

The study was in accordance with the “Principles of laboratory animal care” (NIH publication No. 85-23, revised 1985), and was approved by the Institution’s Animal Care and Use Committee of our University (protocol number 202/2011).

Biometric Measures

Body weights were measured weekly (Marte A-500®). Additionally, at the end of the 8th week animals were anesthetized and sacrificed by cervical dislocation. The hearts were quickly removed and the heart weight was measured (Marte A-500®). Finally, the index of cardiac hypertrophy was expressed as quotient between heart and body weights.

Electrocardiogram Acquisition and Measurement

Prior to ECG recordings, animals were conditioned for 7 consecutive days, 20 minutes each day, inside a plexiglass restrainer. All posterior recordings were conducted in a constant environment, during the morning (0700-1100h).

After carefully shaving the ventral thoracic region of animals, they were clothed with a custom-made elastic cotton jacket developed to fit the rat’s mean thoracic circumference. Two rectangular pieces of platinum electrodes (7.0 x 3.0 mm) were attached to the jacket’s inner surface. The ECG was acquired in a lead close to DII, maintaining prominent R wave peaks. A conductive ECG gel was applied over each electrode, with care being taken to avoid the establishment of a gel bridge between them. Animals were placed inside a plexiglass restrainer (Figure 1). More details of ECG acquire method see [29]. Electrodes were connected to a differential A/C amplifier (A-M Systems, USA), digitized by a 16 bit A/D interface converter (Digidata 1322-A, Axon Instruments, USA), and sampled at 2 KHz by the software Axoscope 9.0 (Axon Instruments, USA). Data were stored in a PC for off-line processing. The ECG recording started 5 minutes after placing each animal inside the plexiglass restrainer and was conducted for 12 minutes [29].

Illustrates the validated method for electrocardiogram acquisition in conscious rats. Jacket placement on the rat (A). Schematic diagram of the inner face of the jacket, illustrating negative and positive electrodes positions. LAL – left anterior limb; RAL – right anterior limb (B). Restrained animal in rest during signal acquisition (C). (Adapted from Annals of the Brazilian Academy of Sciences²⁹)

Heart Rate Variability Analysis

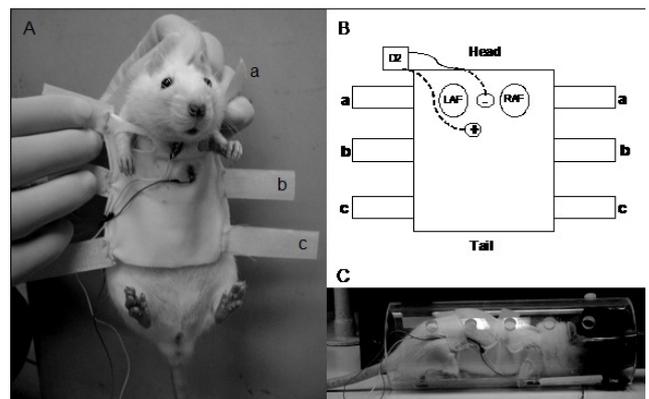


Figure 1. Electrocardiogram acquisition method

Firstly, from the ECG signal, the beat-to-beat R-R intervals were extracted using the Pan-Tompkins algorithm [30]. Furthermore, 180-s tachograms were generated, containing all heart period fluctuations within this time segment and the series of artifact were de-emphasized using a variable threshold R wave filter in accordance to the criteria of Berntson et al. (1990) [31]. The following indexes were extracted in the time-domain: RR (mean RR interval), SDNN (standard deviation of RR intervals), RMSSD (square root of the mean squared differences of successive RR intervals) and pNN5 (percentage of successive RR interval differences greater than 5 ms)[32]. For spectral (frequency-domain) analysis of HRV, tachograms were resampled to equal intervals by spline cubic interpolation method at 10 Hz, and the linear trend was removed. The tachogram signal was divided into segments of 256 intervals

overlapping each other by half. After the removal of the linear trend and application of the Hamming window, each segment was padded with 256 zeros, submitted to a fast Fourier transform (FFT), and magnitude-squared for calculation of the power spectrum according to the periodogram method. The power spectra of all segments were averaged to reduce the variance of FFT as a spectral estimator. Moreover, the obtained average power spectrum was smoothed using a three point sliding rectangular window. According to previous HRV studies in rats [25, 33,34], two regions of interest were defined: low-frequency (LF; >0.2 Hz, <0.8 Hz) and high-frequency bands (HF; ≥ 0.8 Hz up to Nyquist frequency, determined by the mean RRi of the tachogram), expressed as percent of total spectral power.

Additionally, the Poincaré analysis was applied for each tachogram. As previously described [35, 36] this method consists in plotting a vector ($RR_1, RR_2, \dots, RR_{N-1}$) called RR_i as a function of RR_{i-1} (RR_2, RR_3, \dots, RR_N). Sequentially, a number of descriptors were obtained: SD2 (the standard deviation of the projection of the Poincaré plot on the line of identity) and SD1 (the SD of projection of the Poincaré plot on the line perpendicular to the line of identity) representing geometrically the rotation of RR_1 vs RR_{i-1} . As widely accepted, SD1 reflects the short-term variability, and SD2, both short-term and long-term variability. Furthermore, the total variability was represented in the Poincaré plot by the area of the ellipse (s) of radius SD1 and SD2 (eq. 1):

$$s = \pi \cdot SD1 \cdot SD2 \quad (1)$$

Statistics

Data was expressed as mean \pm standard error of the mean (SEM). Each spectral as well as temporal parameter was compared within groups (DECA and Control) and throughout the eight weeks. Two way ANOVA and post hoc Tukey test was used for repeated measures. Signals were processed in Matlab v 6.5 release 13 (Mathworks, USA) and the statistical procedures were made in Statistica v 6.0 (Statsoft, USA). Statistical significance was established at the $P < 0.05$ level.

3. Results

The effects of AAS treatment on the body weight of rats

showed no differences. The final heart weight was not different between the groups when compared in absolute and relative values (Table 1).

Figure 1 illustrates the mean power spectrum in DG throughout the eight weeks of AAS treatment. As depicted by panels A to H, there is a reduction in the PSD total area regarding all the frequency components. During the first week the total PSD averaged $81.9 \pm 15.7 \text{ ms}^2$, reaching $41.0 \pm 6.4 \text{ ms}^2$ at the end of the 8th week ($P < 0.05$), while in the vehicle, the total PSD established around $51.8 \pm 14.3 \text{ ms}^2$.

Nonetheless, this characteristic was quite stronger in the high frequency component, and further, is noticeable in Figure 2 (Panel A), where the PSD for the HF component is plotted. Although only significant differences have been found between DG and CG groups during the fourth ($P = 0.01$), sixth ($P = 0.02$) and eighth ($P = 0.006$) weeks, the tendency to reduce the HF component is supported by the differences achieved along time only for the DECA group, from the 4th until the 8th week ($P < 0.05$). Besides the decreasing in the HF PSD% of the vehicle throughout treatment, the DG decreased from 24.5 ± 7.2 to 3.9 ± 2.5 ($P < 0.01$), corresponding to an alteration of 83.9%. Moreover, given that the total PSD diminished in both groups, the relative PSD for HF (Figure 2 A) corresponded to $26.3 \pm 8.3\%$ and $26.5 \pm 4.1\%$ at the first and eighth weeks (n.s.) for vehicle, whereas the DG reduced significantly from $26.4 \pm 3.6\%$ (1st week) to $18.9 \pm 2.6\%$ (8th week) ($P = 0.031$). Accordingly, panel B (Figure 2) shows a progressive increasing in the LF-HF ratio quite expressive in the group treated with AAS resulting in significant increasing from the 6th to the 8th week, and similarly differences between groups in the 4th ($P = 0.03$), 6th ($P = 0.02$), 7th ($P = 0.04$) and 8th ($P = 0.002$) weeks.

Figure 3 illustrates the Poincaré plot for both groups. The top panels represent the CG (A) and DG (B) in the first week, and the bottom panels are CG (C) and DG (D) in the eighth week. In view of the above considerations, it is possible to distinguish a reduction in the orthogonal dispersion to the identity line, corresponding to a flattening in the ellipsis. This effect can be represented by the SD1 and further corresponds to the short-term variability and likewise the high frequency component of PSD.

Table 1. Effect of nandrolone Decanoate on body and heart weights of male rats

	Body Weight (G)		Final Heart Weight	
	Initial	Final	Absolute (G)	Relative ($\text{Mg} \cdot \text{G}^{-1}$)
Control	275.75 ± 1.40	323.25 ± 14.33	1.97 ± 0.0272	6.06 ± 0.263
Deca	276.10 ± 10.97	303.90 ± 10.86	2.05 ± 0.0507	6.62 ± 0.229

Data expressed as mean \pm SEM. Relative final heart weight: ratio of absolute final heart weight to final body weight * $P < 0.01$, ** $P < 0.001$, versus CONTROL

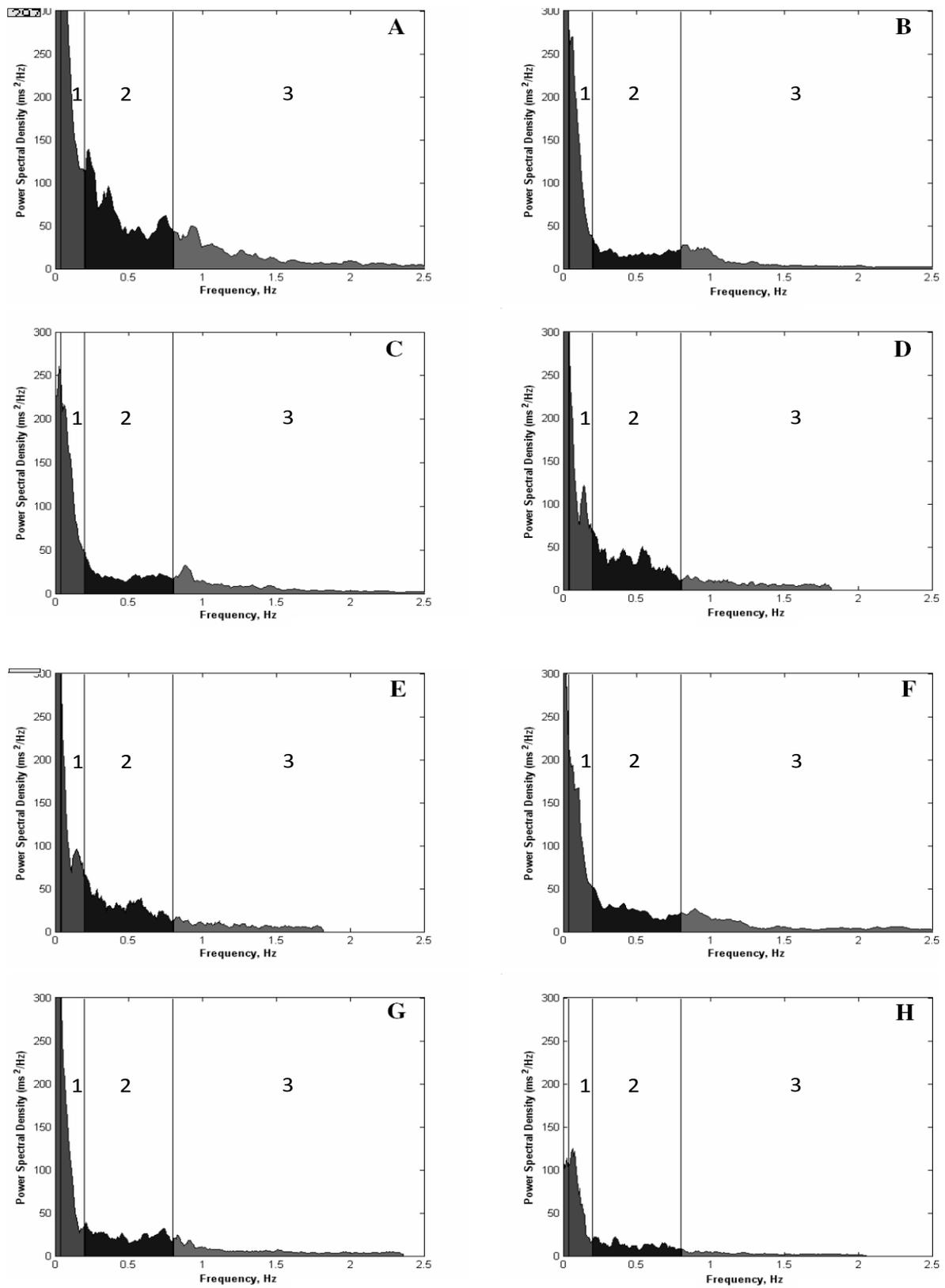


Figure 2. Power Spectral Density of DECA throughout the eight weeks. A, first week; B, second week; C, third week; D, fourth; E, fifth; F, sixth; G, seventh; H, eighth. Area filled with dark gray (1 area) represents the very low frequency component, black (2 area), the low frequency component and light grey (3 area), the high frequency component

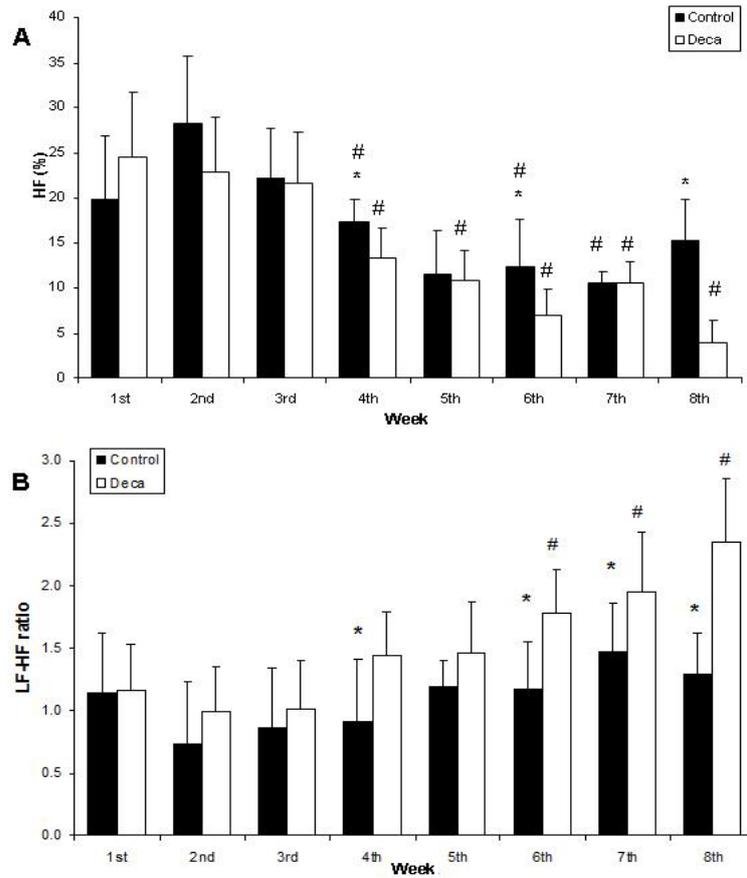


Figure 3. Time course of cardiac autonomic activity throughout the eight weeks of treatment with nandrolonedecanoate. Time course of high frequency (HF) component (A) and the LF-HF ratio (B) throughout the eight weeks of treatment with nandrolonedecanoate (DG) or vehicle (CG); * ($P < 0.05$) represents CG vs. DG in each week and # ($P < 0.05$) represents intragroup comparison vs 1st week

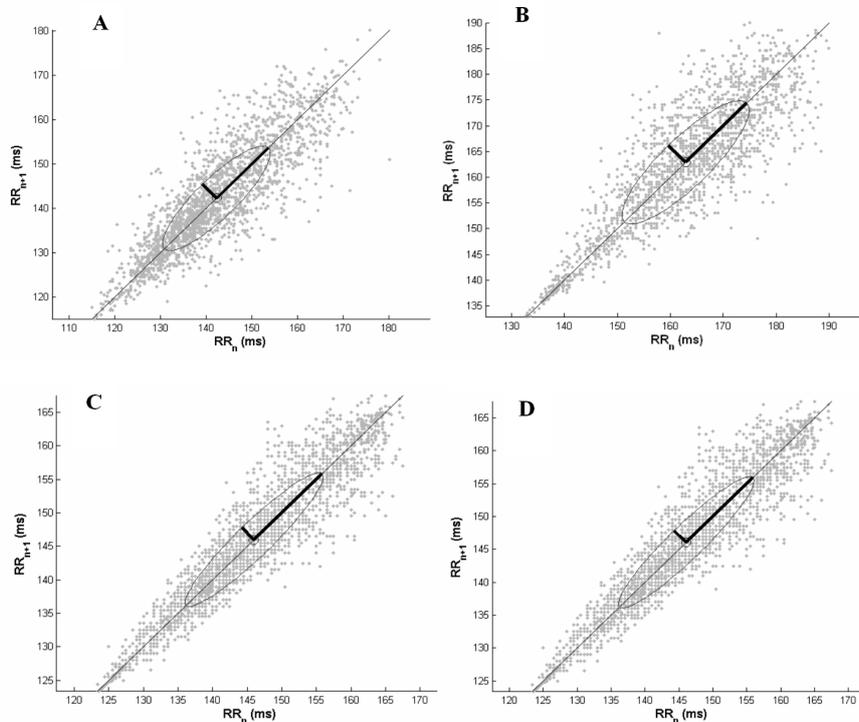


Figure 4. Poincaré plot of typical animals. Left panels represent vehicle group (CG) in the first (A) and eighth (C) week, and the right panels, nandrolonedecanoate group (DG) in the first (B) and eighth (D) week

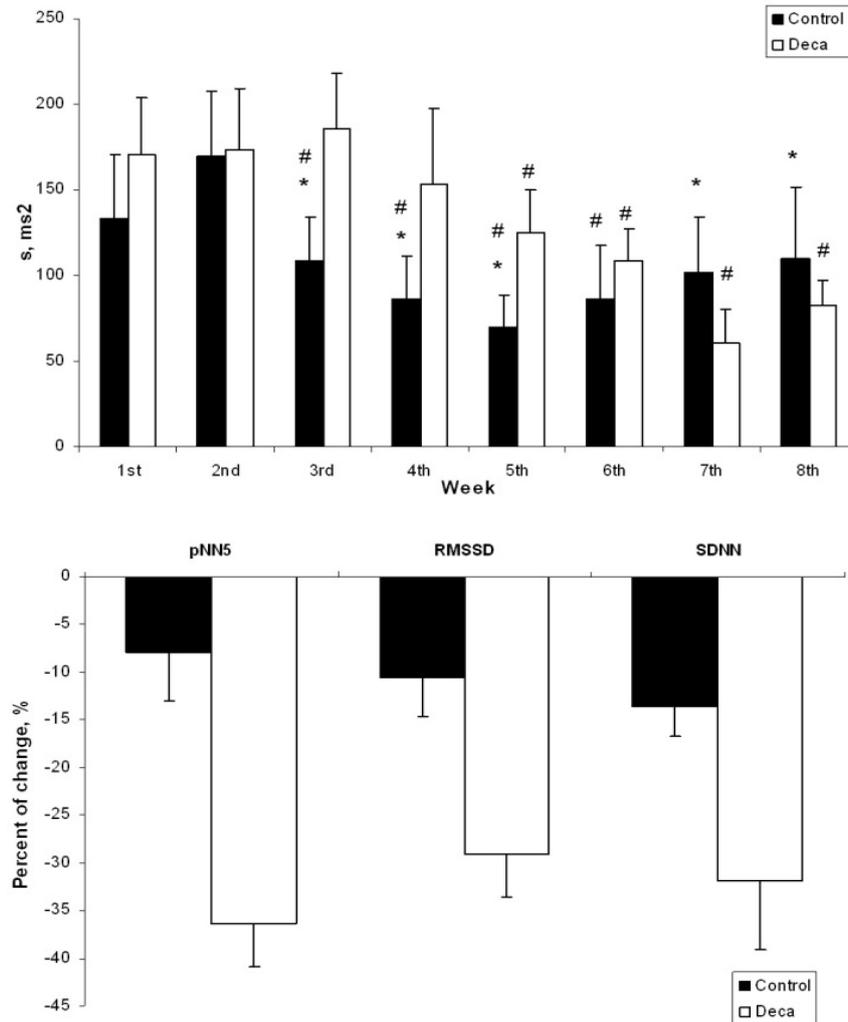


Figure 5. Area (s) from Poincaré Plot throughout the eight weeks in both DG and CG groups and percent of change in time-domain indexes of heart rate variability after the eight weeks in both DG and CG groups. Time course of the Area (s) obtained from the Poincaré plot throughout the eight weeks treatment with nandrolonedecanoate (DG) or vehicle (CG). * ($P < 0.05$) represents CG vs. DG in each week and # ($P < 0.05$) represents intragroup comparison vs 1st week (A). Percent of change in pNN5, RMSSD and SDRR after the eight weeks treatment with nandrolonedecanoate (DG) or vehicle (CG) (B)

Accordingly, good correlation coefficients were found between SD1 and HF averaging 0.75 (DG) and 0.69 (CG). The ellipsis area (s) formed by the two deviations (SD1 and SD2) according to Equation 1 is plotted in Figure 4A. Even though both groups tended to present a reduction in s, this finding is noticeable in the DG, presenting a significant reduction from the 5th week. In accordance to the reduction in the total PSD, the ellipsis area decreased similarly in both groups ($r^2 = 0.83$).

In the time-domain analysis, RMSSD, pNN5 and the SDNN were significantly reduced in the AAS treated group. As showed in Figure 4B, decreasing of these markers decreased 30 to 36% ($P < 0.001$) throughout the eight weeks of AAS treatment.

4. Discussion

To our knowledge, this is the first report to evaluating the cardiac autonomic control assessed weekly following two

different methods based exclusively on the HRV analysis. We showed that the time-course of deleterious effects over the heart function were able to detect early signals of cardiac autonomic dysfunction in rats chronically treated with DECA. Furthermore, the noninvasive method for ECG acquisition was of remarkable use, since it allowed a continuous evaluation of HRV over the 8-week period.

Cardiac hypertrophy is often associated to sudden death and arrhythmias in endurance athletes and AAS users[37-39] and further, a number of autonomic dysfunctions may be generated by the cardiac remodeling[40]. In rats, the degree of cardiac hypertrophy has been accessed through the heart weight/body weight ratio[41]. Accordingly, Andrade et al. [42] showed an increase in heart weight and heart weight/body weight ratio after 4-weeks of DECA treatment in rats, confirming the left ventricular hypertrophy by morphometrical and histological analysis. Even though the results achieved here may corroborate the well-known AAS-induced effect on body weight gain of rats, contrarily to previous works[25], in the present study no significant

increase in the weight/body weight ratio was recorded. It is possible that this divergence may have occurred due to differences in animal weight at the start of treatment, once Pereira-Jr [25] used animals with ~ 357 g. Different from our previous findings [25] in which older animals were treated with DECA, resulting in significant attenuation of body weight gain, herein no significant differences in the animal body weight were observed between groups.

Some investigations showed that recognized disturbances in “core” patterns, indicating progressive destabilization of cardiac rhythm, which would predict the onset of spontaneous sustained ventricular tachyarrhythmias. The possible concomitance of both vagal and sympathetic actions, most likely on a reflex basis, could facilitate arrhythmias by a complex interplay [43]. In fact, the identification of cardiac autonomic dysfunction is frequently used as predictor of sudden death and arrhythmias [20, 24]. Pereira-Junior *et al.* [25] showed alterations in parasympathetic cardiac autonomic modulation only after 8-week treatment AAS treatment. However, our results demonstrated that cardiac autonomic dysfunction may occur earlier, with significant reduction in the HF PSD and increase in the LF/HF ratio in the DG, starting by the 4th week of treatment. In this regard, it may not entirely discard the hypothesis of the involvement of an acute, non-genomic effect of DECA taking place in the genesis of arrhythmia triggering factors. In fact, Phillis and colleagues [44] recently demonstrated that DECA acutely potentiates the arrhythmogenic effect of cardiac ischemia in rats, decreasing animal survival during an ischemic event. Furthermore, it has been recently shown [45] that Clenbuterol, a beta 2-agonist, which is also misused for increasing muscle mass, may lead to acute arrhythmic events. However, according to Andrade *et al.* [42] no alterations in the Bezold-Jarisch reflex were found after 4 weeks of treatment with DECA. In face of these findings, we can suggest that the parasympathetic autonomic dysfunction showed in the present work, starting by the fourth week of DECA treatment, may be a possible result of central abnormalities in cardiovascular autonomic control rather than alterations in cardiac receptor activities. These results suggest that DECA treatment creates a conspicuous substrate to induce electrical disturbances. [27, 28]

In addition, it is well established that Poincaré plot analysis allow visual information about heart rate successive fluctuation patterns, constituting a valuable and simple method of evaluating cardiac autonomic control [26]. This method is advantageous in some ways because it constitutes a simple tool that requires low computational cost in order to represent similar information to that extracted from spectral analysis, without a need for stationary and periodic signals for analysis computation. We showed that DG tended to present a significant reduction in s , from 5th week, and the ellipsis area decreased similarly in both groups correlating with the reduction in the total PSD ($r^2 = 0.83$). However, the time-course of s variation was different between groups, with a marked decrease from 5th to 8th week in DG while CG

presented preserved pattern of autonomic modulation by the end of treatment.

In the present work, the time-domain HRV variables RMSSD and pNN5 were lower in AAS group compared to the control group. The lower values of pNN5 in the AAS group suggest parasympathetic cardiac dysfunction [Maioreschi, 2012 #46]. However, the RMSSD index has better statistical properties for assessing parasympathetic activity than pNN5 [Berntson, 1996 #52]. The RMSSD has been used as a better index of cardiac parasympathetic control since it is uncontaminated by sympathetically mediated HRV [Berntson, 2005 #56] and suggesting an impairment in the parasympathetic reactivation in rats treated with supraphysiological doses of nandrolonedecanoate. The SDNN value in DECA group was lower compared to the control group. Some studies have associated the decrease of SDNN value with left ventricular dysfunction and development of high blood pressure [Casolo, 1992 #58] [Schroeder, 2003 #61]. Thus, supraphysiological doses of AAS seem to induce an autonomic dysfunction that reflects in a reduction of the variability of R-R intervals and in a lower SDNN [Maioreschi, 2012 #46].

The mechanism underlying the cardiac autonomic dysfunction in AAS-abusing remains unknown. However, Previous studies have reported that androgens can cross the blood-brain barrier [Kindlundh, 2004 #62] [Kicman, 2008 #67] and bind to androgen receptors in different brain regions [Sheridan, 1982 #69] [Simerly, 1990 #70] [Penatti, 2009 #72]. Androgens may also be aromatized to estrogens and bind to estrogen receptors [Penatti, 2009 #72]. AAS affect GABAergic transmission in the hypothalamus and other brain areas by modulation of GABA_A receptor function [Henderson, 2006 #76]. Thus, one might speculate that AAS alter the parasympathetic modulation mediated by central GABA and other mechanisms, by binding to androgen receptors in the hypothalamus and brain stem regions that control the tonic and reflex response of cardiovascular system.

5. Conclusions

In conclusion, we showed that cardiac autonomic dysfunction may constitute an early consequence of the chronic administration of high doses of anabolic steroids. Besides, using a noninvasive approach for assessing heart rate variability in rats and Poincaré analysis method, we have demonstrated that animals treated with nandrolonedecanoate show important reductions in parasympathetic and increases in cardiac sympathetic modulation, which may constitute an important marker of arrhythmia vulnerability and sudden death identification.

List of abbreviations

AAS, Anabolic androgenic steroids; CG, Control group treated with vehicle; DECA, nandrolonedecanoate; DG, Group treated with nandrolonedecanoate; ECG, Electrocardiogram; FFT, Fast Fourier transform; HF, High

frequency component of heart rate variability; HRV, heart rate variability; LF, Low frequency component of heart rate variability; PNN5, Percentage of successive RR interval differences greater than 5 ms; PSD, Power spectral density; RMSSD, Square root of the mean squared differences of successive RR intervals; RR, mean of RR interval of electrocardiogram; SDNN, standard deviations of RR intervals; SD1, standard deviation of projection of the Poincaré plot on the line perpendicular to the line of identity; SD2, the standard deviation of the projection of the Poincaré plot on the line of identity.

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